

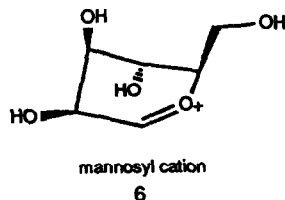
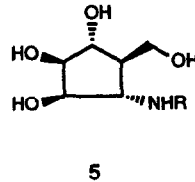
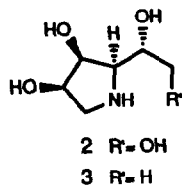
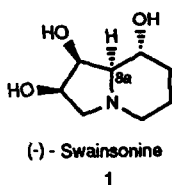
SYNTHESIS OF 15,2R,3S,4R,5R-METHYL[2,3,4-TRIHYDROXY-5-(HYDROXYMETHYL)CYCLOPENTYL]AMINE: A POTENT α -MANNOSIDASE INHIBITOR

Robert A. Farr,* Norton P. Peet, and Mohinder S. Kang

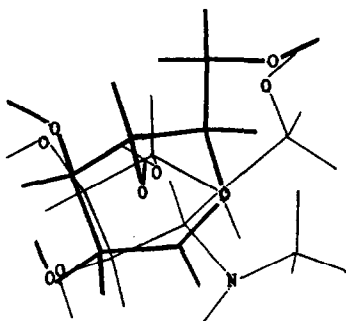
Merrell Dow Research Institute
2110 East Galbraith Road
Cincinnati, OH 45215

Summary: The title carbocyclic amine **15**, prepared in high yield from isoxazolidine **8** using an efficient oxidation-epimerization-reduction sequence, is an inhibitor of mung bean α -mannosidase II and a potent inhibitor of jack bean α -mannosidase.

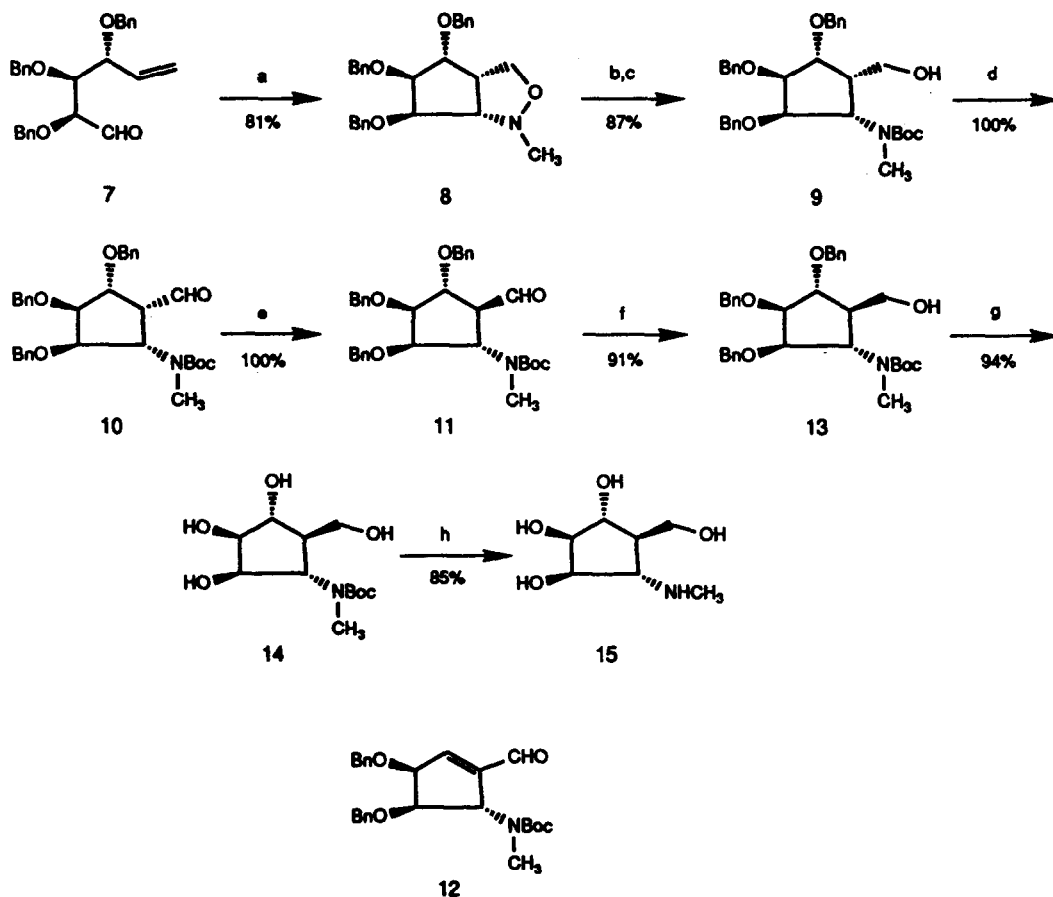
Inhibitors of the glycoprotein processing enzyme α -D-mannosidase II, such as swainsonine (**1**),¹ have potential therapeutic use as antimetastatic,^{2,3} and immunomodulatory agents.⁴ Synthetic hydroxylated pyrrolidines such as 1,4-dideoxy- and 1,4,6-trideoxy-1,4-imino-D-mannitol (**2**⁵ and **3**⁶) have shown potent α -mannosidase inhibitory activity. A recent molecular orbital and molecular graphics study⁷ concluded that **1-3**, as well as **8a-epi 1** are more potent α -mannosidase inhibitors than the more mannose-like deoxymannojirimycin (**4**) because of a better superimposition of their heteroatoms with the corresponding heteroatoms of the lowest energy "flap up" half chair form of the mannosyl cation **6**,⁷ the presumed



intermediate in the hydrolysis of mannopyranosides. We proposed the hydroxylated cyclopentylamines 5 as potential α -mannosidase inhibitors based on the following: a) the α -amino function is appropriately positioned to be protonated in the enzyme active site by an α -oriented carboxylic acid, and b) four hydroxy groups, which are ideally oriented because of the geometry of the five-membered ring, are present, as in the mannosyl cation 6. Molecular modeling studies of N-methyl analog 15 vs the MO-optimized "flap up" half chair form of the mannopyranosyl cation 6⁷ show good overlap of the hydroxy groups. The N-methylamino moiety is located on the α -face between the ring oxygen and C-1 of the mannosyl cation, with a closer proximity to C-1. Consequently, cyclopentylamines 5 could mimic either the mannosyl cation or its protonated polysaccharide precursor.

cyclopentylamine 15mannosyl cation 6

The known isoxazolidine 8⁹ was prepared in 81% yield via the intramolecular cycloaddition of the N-methyl nitron derived from aldehyde 7. Reductive cleavage of the isoxazolidine ring (Zn dust, aq 85% HOAc, 50-55°C, 1 h) followed by acylation of the crude amino alcohol [(BOC)₂O, THF, 50°C] gave α -hydroxymethyl carbamate 9.¹⁰ Swern oxidation [2.7 equiv DMSO, 2.3 equiv (COCl)₂, 5.1 equiv DBU, -78°C] gave a mixture of products, including 43% of the epimerized β -carboxaldehyde 11 and 10% of the α,β -unsaturated aldehyde 12; no 10 was detected. However, oxidation of 9 with the Dess-Martin periodinane¹¹ (1.5 equiv, CH₂Cl₂, 30 min) cleanly gave the α -carboxaldehyde 10 in quantitative yield (crude). Treatment of 10 with 0.25 equiv DBU at -78°C (CH₂Cl₂, 1 h, quench with HOAc at -78°C) led to clean epimerization to 11 without any concomitant elimination. Reduction of 11 with NaBH₄ in EtOH gave the β -hydroxymethyl carbamate 13 in 91% overall yield from 9. Debenzylation (H₂, Pd black, CH₃OH, 4 days) to 14 followed by cleavage of the t-Boc group [HCl(g), ether/CH₃OH] and flash chromatography¹² on silica gel (3:1:2 CH₃OH:conc NH₄OH:CH₂Cl₂) gave 15. Further purification by ion exchange chromatography (Bio-Rad AG 50W-X8, 0.5 M NH₄OH) and crystallization from *i*-PrOH at -78°C gave 15 as an extremely hygroscopic white solid.



(a) CH_3NHOH , CH_3OH , Δ ; (b) Zn , HOAc ; (c) $(\text{Boc})_2\text{O}$; (d) Dess-Martin periodinane; (e) DBU , -78°C ; (f) NaBH_4 ; (g) H_2 , Pd ; (h) HCl(g) .

The carbocyclic amine **15** is a potent inhibitor of jack bean α -mannosidase,¹³ with an IC_{50} of 62 nM vs an IC_{50} of 87 nM for swainsonine (**1**). However, against the purified mung bean glycoprotein processing enzyme α -mannosidase II¹⁴, **15** has an IC_{50} of 1.0 μM as compared to swainsonine which has an IC_{50} of 40 nM. The potent activity of **15** is exceptional in that recent reports show that *N*-methylation of monocyclic analogues of swainsonine markedly decreases α -mannosidase inhibitory activity.¹⁵

Acknowledgments

We thank Dr. Herschel Weintraub for modeling the mannosyl cation.

References and Notes

- Swainsonine and Related Glycosidase Inhibitors; James, L.F.; Elbein, A.D.; Molyneux, R.J.; Warren, C.D., Ed.; Iowa State University Press: Ames, 1989.
- (a) Humphries, M.J.; Matsumoto, K.; White, S.L.; Olden, K. *Proc. Natl. Acad. Sci. U.S.A.* **1986**, *83*, 1752. (b) Humphries, M.J.; Matsumoto, K.; White, S.L.; Molyneux, R.J.; Olden, K. *Cancer Res.* **1988**, *48*, 1410.
- Dennis, J.W. *Cancer Res.* **1986**, *46*, 5131.
- (a) Hino, M.; Nakayama, O.; Tsurumi, Y.; Adachi, K.; Shibata, T.; Terano, H.; Kohsaka, M.; Aoki, H.; Imanaka, H. *J. Antibiot.* **1985**, *38*, 926. (b) Kino, T.; Inamura, N.; Nakahara, K.; Kiyoto, S.; Goto, T.; Terano, H.; Kohsaka, M.; Aoki, H.; Imanaka, H. *Ibid.* **1985**, *38*, 936.
- (a) Fleet, G.W.J.; Smith, P.W.; Evans, S.V.; Fellows, L.E. *J. Chem. Soc., Chem. Commun.* **1984**, 1240. (b) Palamarczyk, G.; Mitchell, M.; Smith, P.W.; Fleet, G.W.J.; Elbein, A.D. *Arch. Biochem. Biophys.* **1985**, *243*, 35. (c) Daniel, P.F.; Newburg, D.S.; O'Neil, N.E.; Smith, P.W.; Fleet, G.W.J. *Glycoconjugate J.* **1989**, *6*, 229. (d) Cenci di Bello, I.; Fleet, G.W.J.; Namgoong, S.K.; Tadano, K.-I.; Winchester, B. *Biochem. J.* **1989**, *259*, 855.
- Eis, M.J.; Rule, C.J.; Wurzburg, B.A.; Ganem, B. *Tetrahedron Lett.* **1985**, *26*, 5397.
- Winkler, D.A.; Holan, G. *J. Med. Chem.* **1989**, *32*, 2084.
- (a) Leaback, D.H. *Biochem. Biophys. Res. Commun.* **1968**, *32*, 1025. (b) Cogoli, A.; Semenza, G. *J. Biol. Chem.* **1975**, *250*, 7802. (c) Dorling, P.R.; Huxtable, C.R.; Colegate, S.M. *Biochem. J.* **1980**, *191*, 649.
- Bernet, B.; Vasella, A. *Helv. Chim. Acta* **1979**, *62*, 2400.
- Physical data for new compounds: **9**, white solid: mp 74.5-77°C; IR (KBr) ν_{\max} 3512, 1682 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3) δ 7.4-7.2 (m, 15 H), 4.72-4.42 (m, 7 H), 4.52 (d, $J=12$ Hz), 4.44 (d, $J=12$ Hz), 4.32-4.17 (m, 1 H), 4.01 (d, 0.5 H, $J=2$ Hz), 3.99 (d, 0.5 H, $J=2$ Hz), 3.96-3.86 (m, 1 H), 3.63-3.54 (m, 2 H), 2.8 (m, 1 H), 2.79 and 2.74 (2s, 3 H), 2.63 (m, 0.5 H), 2.17 (m, 0.5 H), 1.47 (s, 9 H); mass spectrum, m/z 548 (M^+), 448 (100); $[\alpha]_D^{25} + 56^\circ$ (c 0.27, CHCl_3). Anal. ($\text{C}_{33}\text{H}_{41}\text{NO}_6$) C, H, N. **10**, colorless oil: $^1\text{H NMR}$ (CDCl_3) δ 9.68 (bs, 1 H), 7.4-7.1 (m, 15 H), 4.75-4.35 (m, 8 H), 4.19-3.86 (m, 2 H), 3.30 (bs, 1 H), 2.78 and 2.72 (2s, 3 H), 1.45 (s, 9 H). **11**, colorless oil: $^1\text{H NMR}$ (CDCl_3) δ 9.69 (s, 1 H), 7.4-7.23 (m, 15 H), 4.86 (t, 1 H, $J=8.4$ Hz), 4.6-4.4 (m, 6 H), 4.15-4.05 (m, 2 H), 3.87 (bs, 1 H), 2.74 (bs, 4 H), 1.45 (s, 9 H). **12**, $^1\text{H NMR}$ (CDCl_3) δ 9.75 (s, 1 H), 7.4-7.28 (m, 10 H), 6.79 (bs, 1 H), 5.18 (bs, 0.4 H), 4.85 (d, 0.6 H, $J=1.0$ Hz), 4.80-4.52 (m, 6 H), 4.23 and 4.07 (2 m in 3:2 ratio, 1 H), 2.80 and 2.67 (2s in 3:2 ratio, 3 H), 1.43 (s, 9 H). **13**, colorless oil: IR (neat) ν_{\max} 3458, 1692, 1668 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3) δ 7.38-7.25 (m, 15 H), 4.65-4.37 (m, 7 H), 3.98 (dd, 1 H, $J=8.9, 5$ Hz), 3.84 (dd, 1 H, $J=5, 2.5$ Hz), 3.81-3.54 (m, 4 H), 2.70 and 2.64 (2 s in 1:2.5 ratio, 3 H), 1.98 and 1.88 (2 bs in 3:8 ratio, 1 H), 1.48 (s, 9 H); mass spectrum, m/z 548 (M^+), 492 (100); $[\alpha]_D^{25} + 44.8^\circ$ (c 1.02, CHCl_3). Anal. ($\text{C}_{33}\text{H}_{41}\text{NO}_6$) C, H, N. **14**, white crystals: mp 81-83°C; IR (KBr) ν_{\max} 3420, 1690, 1666 cm^{-1} ; $^1\text{H NMR}$ (CDCl_3) δ 5.22 (bd, 2 H, $J=5.6$ Hz), 4.64 (bs, 1 H), 4.58 (bs, 1 H), 4.15 (bs, 2 H), 3.94 (bs, 1 H), 3.73 (bs, 1 H), 3.54 (bs, 1 H), 2.81 (s, 3 H), 1.78 (bs, 1 H), 1.44 (s, 9 H); FABMS (glycerol), m/z 278 (M^+), 222 (100); $[\alpha]_D^{25} + 30.8^\circ$ (c 1.06, CHCl_3). Anal. ($\text{C}_{12}\text{H}_{23}\text{NO}_6$) C, H, N. **15**: $^1\text{H NMR}$ (D_2O) δ 3.86-3.59 (m, 5 H), 2.61 (dd, 1 H, $J=7.3, 2.9$ Hz), 2.31 (s, 3 H), 1.55 (m, 1 H); $^{13}\text{C NMR}$ (D_2O) δ 79.34, 78.16, 75.94, 67.98, 64.28, 52.10, 35.92; $[\alpha]_D^{20} + 8.5^\circ$ (c 1.07, CH_3OH). Anal. ($\text{C}_8\text{H}_{15}\text{NO}_4$) H, N; C: calcd, 47.45; found 46.58.
- Dess, D.B.; Martin, J.C. *J. Org. Chem.* **1983**, *48*, 4155.
- Still, W.C.; Kahn, M.; Mitra, A. *J. Org. Chem.* **1978**, *43*, 2923.
- Kang, M.S.; Elbein, A.D. *Plant Physiol.* **1983**, *71*, 551.
- Kaushal, G.P.; Elbein, A.D. In *Methods Enz.*; Gingsburg, V., Ed.; Academic Press, Inc.: San Diego, 1989; Vol. 179, pp 468-471.
- (a) Al Daher, S.; Fleet, G.; Namgoong, S.K.; Winchester, B. *Biochem. J.* **1989**, *258*, 613. (b) Schweden, J.; Legler, G.; Bause, E. *Eur. J. Biochem.* **1986**, *157*, 563.